



## Review

### Pharmacological classification of herbal anti-asthmatics

Bhoomika R Goyal<sup>1,2</sup>, Babita B Agrawal<sup>2</sup>, Ramesh K Goyal<sup>2</sup> and Anita A Mehta<sup>2,\*</sup>

<sup>1</sup>Institute of Pharmacy, Nirma University of Science and Technology, Ahmedabad - 382 481, Gujarat, India;

<sup>2</sup>Department of Pharmacology, L. M. College of Pharmacy, Ahmedabad - 380 009, Gujarat, India

#### SUMMARY

Bronchial asthma is a major public health problem worldwide and the morbidity and mortality of asthma have increased in last few decades. There is high prevalence of usage of alternative traditional system of medicines for the treatment of asthma. Large numbers of medicinal plant preparations have been reported to possess anti-asthmatic effects. Plant cells are now considered to be the chemical factories synthesizing a large variety of chemical compounds. Further, Ayurvedic system of medicine has an elaborate description of asthma from the earliest times describing it as '*Shwasa*' meaning disease pertaining to breathing. This review classifies the anti-asthmatics herbs based on the possible mechanism of action reported. Thus, these plants can be used to obtain a polyherbal formulation which contains various herbs acting at particular sites of the pathophysiological cascade of asthma for prophylaxis as well as for the treatment of asthma.

**Key words:** Anti-asthmatics herbs; Asthma

#### INTRODUCTION

Primary respiratory diseases are responsible for a major burden of morbidity and ultimately deaths and lungs are often affected in multi-system diseases. Bronchial asthma is a major public health problem worldwide and the morbidity and mortality of asthma have increased in last few decades. The past decade has witnessed phenomenal increases in the incidences of asthma, asthma-related deaths and hospitalization. An estimated 12 million persons in United States have asthma. India has an estimated 40 million asthmatics (WHO Fact Sheet, 2000). The activation of cells bearing allergen-

specific IgE initiates the *early phase reaction*. It is characterized primarily by the rapid activation of airway mast cells and macrophages. The activated cells rapidly release pro-inflammatory mediators such as histamine, eicosanoids and reactive oxygen species that induce contraction of airway smooth muscle, mucus secretion and vasodilatation. Inflammatory mediators induce microvascular leakage with exudation of plasma in the airways. Together these effects contribute to airflow obstruction. The second, *late-phase response*, i.e. the delayed response, occurs 6 to 9 h after allergen provocation and involves the recruitment and activation of eosinophils, CD4<sup>+</sup> T cells, basophils, neutrophils and macrophages. The activated T-lymphocytes also direct the release of inflammatory mediators from eosinophils, mast cells and lymphocytes. In addition, the subclass 2 helper T-lymphocytes

\*Correspondence: Anita A Mehta, Department of Pharmacology, L. M. College of Pharmacy, Ahmedabad - 380 009, Gujarat, India. Tel: +91-79-2630-2746; Fax: +91-79-2630-4865; E-mail: dranitalmcp@rediffmail.com

subset of activated T-lymphocytes produces interleukin (IL)-4, IL-5 and IL-13. IL-4 in conjunction with IL-13 signals the switch from IgM to IgE antibodies. IL-5 activates the recruitment and activation of eosinophils. This phase is in essence a progressing inflammatory reaction. Regardless of the triggers of asthma, the repeated cycles of inflammation in the lungs with injury to the pulmonary tissues followed by repair may produce long-term structural changes (“remodeling”) of the airways (Fireman, 2003).

Current pharmacotherapy of asthma comprises the use of bronchodilators (selective  $\beta_2$  agonists, xanthines and anti-cholinergics), anti-inflammatory agents (mast cell stabilizers and corticosteroids), leukotriene antagonists and lipoxygenase inhibitors. Increase in bronchial hyperresponsiveness (Schayck *et al.*, 1990), diminish the duration of bronchoprotective effects is seen with  $\beta_2$  agonists. Xanthines have narrow therapeutic index. Anti-cholinergics like ipatropium incompletely protects against bronchoconstriction induced by histamine, cold air, allergen, exercise, prostaglandins, bradykinin, serotonin and other mediators. Mast cell stabilizers are not indicated for the relief of acute symptoms of asthma as they don't have bronchodilator effects. Long term complications are seen with corticosteroids. Further, none of these agents is able to act at all the stages of asthma and thus do not give complete cure of the disease. As a result, there is high prevalence of usage of alternative traditional system of medicines for the treatment of asthma. Ayurveda offers a unique insight into comprehensive approach to asthma management through proper care of the respiratory tract. Large numbers of medicinal plant preparations have been reported to possess anti-asthmatic effects. Plant cells are now considered to be the chemical factories synthesizing a large variety of chemical compounds. The wide range of structures of the plant constituents, which appear to be the active anti-asthmatic principles, suggests different sites of action within the body. This article is intended to classify anti-asthmatic herbs based on the possible mechanism of action reported.

## INDIAN AYURVEDIC CONCEPT OF ASTHMA

Ayurveda is one of the major traditional medicinal systems from India. The ancient Ayurvedic system of medicine has an elaborate description of this disease from the earliest times. *Shwasa* word in normal terminology means respiration. In the present context, *Shwasa* means disease pertaining to breathing. According to Ayurveda, different types of *Shwasa* (asthma) are *Kshudra Shwasa*, *Maha Shwasa*, *Urdhva Shwasa*, *Chhinna Shwasa*, *Tamak Shwasa*.

### *Kshudra Shwasa*

Because of vitiation of *vayu* (air) in the alimentary tract, minor dyspnoea is caused. This condition does not give much pain; it does not interfere in the course of food and breathing. It does not disturb the sensory organs. This condition is mainly because of excessive intake of *ruksha* eatable and excessive exercise. It is, however, not much harmful to the body as compared to the other types of *Shwasa*. In allopathic system of medicine, such types of conditions are grouped under the exertional dyspnoea. Both the systems of medicine consider this condition to be easily curable.

### *Maha Shwasa*

This condition is caused because of disturbance in respiratory movement of *Vayu*. The patient feels great obstruction in respiration, breaths without break with a very loud and long stertore making a sound like intoxicated bull. The patient loses all senses of understanding and knowledge, having restless look in eyes, distorted voice, going into semi-comatose condition now and then. In such a condition, the patient is not able to pass urine and faces both. In this condition, usually the voluntary control disappears and the wheezing sounds are audible from a distance. The allopathic system of medicine indicates such conditions in Biot's breathing which is generally found in heart, kidney and brain disorders as a complication. Ayurveda describes it

as a dyspnoea major where the patient generally succumbs to it.

#### **Urdhva Shwasa**

Under this condition the expiratory phase is prolonged and the inspiratory process is just insignificant. Mouth and the respiratory tract get obstructed with *Kaph*. The patient's eyes are turned upwards and are restless. The patient is almost oblivious to his surroundings. Affected with severe pain, the patient enters into stupor. Having provoked expiratory process and obstruction in inspiration, the patient suffers from delusions and senselessness. Such condition is described by Ayurvedic physicians as harmful for the life. Such conditions are described by the allopathic system of medicine under stertorous breathing and falling of inspiration. Such a condition can be found in pneumonia, abscesses of the lungs, gangrene or acute inflammation in the lungs and also in different types of epilepsy.

#### **Chhinna Shwasa**

Under this condition, the whole of the breathing system is depressed. The patient has to breathe with full force and with great difficulty. The patient breaths with interruption. The patient suffers from constipation, excessive sweating, repeated fainting, burning and retention of urine, having eyes full of tears and entering unconsciousness every now and then having dry mouth. The patient normally breaks down with such a difficult breath ultimately losing his life. The allopathic system of medicine groups such condition under interrupted respiratory dyspnoea (Cheyne-stoke's respiration).

#### **Tamaka Shwasa**

Acharya classified *Tamak Shwasa* in two conditions viz. *Pratamaka Shwasa* and *Santamak Shwasa*. Febrile dyspnoea appears in a patient with fever and fainting in *Pratamaka Shwasa*. It is excited by misperistalsis, inhalation of dust, indigestion, old age or debilitated condition or the suppression of

natural urges. *Santamak Shwasa* or cardiac asthma is greatly aggravated during night and alleviated by cold medicines and in which the patient feels as if he is submerged in a sea of darkness.

## **ANTI-ASTHMATIC HERBS**

Many Ayurvedic plants have been described to be useful in the treatment of various bronchial disorders including bronchial asthma. The use of medicinal plants and natural products increased dramatically in the last two decades in all over the world. More than 400 medicinal plant species have been used ethnopharmacologically and traditionally to treat the symptoms of asthmatic and allergic disorders worldwide.

### **CLASSIFICATION OF ANTI-ASTHMATIC HERBS BASED ON MECHANISM OF ACTION**

Some herbal alternatives employed in asthma are proven to provide symptomatic relief and assist in the inhibition of disease development as well. These herbs therefore have multifaceted roles to play in the management of asthma suggesting different sites of action within the body. Based on the possible mechanism of action reported, plant anti-asthmatics may be classified as follow.

#### **Bronchodilators**

*Adhatoda vasica*, *Albizia lebbek*, *Artemisia caerulescens*, *Belamcanda chinensis*, *Benincasa hispida*, *Cissampelos sympodialis*, *Clerodendron serratum*, *Coleus forskohlii*, *Elaeocarpus sphericus*, *Galphimia glauca*, *Gardenia latifolia*, *Ginko biloba*, *Ocimum sanctum*, *Passiflora incarnate*, *Pavetta crassipes*, *Picrorrhiza kurroa*, *Sarcostemma brevistigma*, *Tephrosia purpurea*, *Tylophora indica*, *Vitex negundo* (Table 1).

#### **Mast cell stabilizers**

*Achyranthes aspera*, *Albizia lebbek*, *Allium cepa*, *Aquillaria agallocha*, *Azadirachta indica*, *Bacopa monniera*, *Bidens parviflora*, *Calotropis procera*, *Cassia*

**Table 1.** Bronchodilators

Name of plant	Part used/ Extract/Fraction	Major chemical constituent (s)	References
<i>Adhatoda vasica</i>	Leaves, roots	Alkaloids	Paliwa et al, 2000
<i>Albizzia lebbek</i>	Stem bark/ Aqueous	Saponins	Tripathi and Das, 1977
<i>Artemisia caerulea</i>	Aerial parts/ Butanolic	Quercetin, isorhamnetin	Moran et al., 1989
<i>Belamcanda chinensis</i>	Leaves/ Ethanolic	Tectorigenin	Singh and Agrawal, 1990
<i>Benincasa hispida</i>	Fruits/ Methanolic	Triterpenes, glycosides, sterols	Kumar and Ramu, 2002
<i>Cissampelos sympodialis</i>	Leaves and root bark/ Aqueous	Warifteine, $\alpha$ -bisbenzyliso-quinoline alkaloid	Thomas et al., 1995; Thomas et al., 1997; Cortes et al., 1995
<i>Clerodendron serratum</i>	Stem bark/ Aqueous	Phenolic glycoside	Gupta, 1968; Gupta and Tripathi, 1973
<i>Coleus forskohlii</i>	Roots	Forskolin (diterpenoid)	Marone et al., 1987
<i>Elaeocarpus sphaericus</i>	Fruits/ aqueous, pet-ether, benzene, acetone and ethanol	Glycoside, steroids, alkaloid, flavanoids	Singh et al., 2000
<i>Galphimia glauca</i>	Aerial/ alcoholic extract/ ethyl-acetate	Tetragalloylquinic acid, quercetin	Campos et al., 2001
<i>Gardenia latifolia</i>	Bark	Saponins	Gupta, 1974
<i>Ginkgo biloba</i>	Leaves	Ginkgolides	Puglisi et al., 1988
<i>Ocimum sanctum</i>	Leaves/ Ethanolic	Myrcenol, nerol, eugenol	Singh and Agrawal, 1991
<i>Passiflora incarnata</i>	Leaves/ Methanolic		Dhawan et al., 2003
<i>Pavetta crassipes</i>	Leaves/ Aqueous	Flavanoids, tannins, anthraquinones	Amos et al., 1998
<i>Picrorrhiza kurroa</i>	Roots	Androsin	Stuppner et al., 1991, 1993
<i>Sarcostemma brevistigma</i>	Twigs/ Alkaloidal fraction	Bregenin	Saraf and Patwardhan, 1988b
<i>Tephrosia purpurea</i>	Aerial parts/ Ethanolic extract	Flavanoids, tephrosin	Gokhale et al., 2000
<i>Tylophora indica</i>	Leaves/ Alkaloidal fraction	Tylophorine	Nayampalli and Sheth, 1979
<i>Vitex negundo</i>	Leaves/ Ethanolic	Casticin, isoorientin, chrysofenol D, luteolin	Nair and Saraf, 1995

*alata*, *Cassia obtusifolia*, *Cassia torosa*, *Cedrus deodara*, *Citrus unshiu*, *Clerodendron serratum*, *Cnidium monnieri*, *Coleus forskohlii*, *Crinum glaucum*, *Elaeocarpus sphaericus*, *Gleditsia sinensis*, *Impatiens textori*, *Inula racemosa*, *Magnolia officinalis*, *Mentha piperita*, *Ocimum sanctum*, *Picrorrhiza kurroa*, *Siegesbeckia glabrescence*, *Solanum xanthocarpum*, *Striga orobanchioids*, *Tephrosia purpurea*, *Terminalia chebula*, *Tinospora cordifolia*, *Tylophora asthmatica*, *Vitex negundo* (Table 2).

#### Anti-allergic agents

*Adhatoda vasica*, *Albizzia lebbek*, *Alisma orientale*, *Aquillaria agallocha*, *Asiasarum sieboldi*, *Camellia sinensis*, *Centipeda*

*minima*, *Citrus unshiu*, *Cnidium monnieri*, *Crinum glaucum*, *Curcuma longa*, *Dalbergia odorifera*, *Desmodium adscendens*, *Galphimia glauca*, *Ginkgo biloba*, *Gleditsia sinensis*, *Hydrangea macrophylla*, *Inula racemosa*, *Magnolia officinalis*, *Sarcostemma brevistigma*, *Siegesbeckia glabrescence*, *Solanum xanthocarpum*, *Terminalia chebula*, *Vitex negundo* (Table 3).

#### Anti-inflammatory agents

*Asystasia gangetica*, *Calotropis procera*, *Curcuma longa*, *Dalbergia odorifera*, *Elaeocarpus sphaericus*, *Eucalyptus globules*, *Ocimum sanctum*, *Pavetta crassipes*, *Tylophora asthmatica* (Table 4).

**Table 2.** Mast cell stabilizers

Name of plant	Part used/ Extract/Fraction	Major chemical constituent (s)	References
<i>Achyranthes aspera</i>	Aerial parts/ Aqueous	Oleanolic acid	Agrawal <i>et al.</i> , 2003
<i>Albizzia lebeck</i>	Stem bark/ Aqueous	Saponins	Tripathi <i>et al.</i> , 1979
<i>Allium cepa</i>	Bulbs/ Juice	$\alpha$ and $\beta$ unsaturated thiosulphinates	Johri <i>et al.</i> , 1985
<i>Aquillaria agallocha</i>	Stem/ Aqueous extract	Triterpenoids	Kim <i>et al.</i> , 1997
<i>Azadirachta indica</i>	Leaves/ Juice	Nimbin, nimbinine, nimbadiol, quercetin	Acharya <i>et al.</i> , 2003
<i>Bacopa monniera</i>	Leaves/ Ethanolic	Bacosides, alkaloids, glycosides	Samiulla <i>et al.</i> , 2001
<i>Bidens parviflora</i>	Aerial parts	Glycosides	Wang <i>et al.</i> , 2001
<i>Calotropis procera</i>	Latex	$\alpha$ -amyrin, $\beta$ -amyrin calotropin (triterpenoid)	Kumar and Basu, 1994
<i>Cassia alata</i>	Leaves/ Ethanolic	Anthraquinones, flavanoids	Palanichamy <i>et al.</i> , 1991
<i>Cassia obtusifolia</i>	Seeds/ Glycosidal fraction	Anthraquinones, betulinic acid	Kitanaka <i>et al.</i> , 1998
<i>Cassia torosa</i>	Seeds	Gentiobiosides	Kanno <i>et al.</i> , 1999
<i>Cedrus deodara</i>	Wood oil	Himacholol	Shinde <i>et al.</i> , 1999
<i>Citrus unshiu</i>	Peels	Flavanoids	Kim <i>et al.</i> , 1999
<i>Clerodendron serratum</i>	Bark/ Aqueous	Phenolic glycoside	Gupta, 1968
<i>Cnidium monnieri</i>	Fruits/ Ethanolic	Osthol	Chen <i>et al.</i> , 1988
<i>Coleus forskohlii</i>	Roots	Forskolin (diterpenoid)	Marone <i>et al.</i> , 1987
<i>Crinum glaucum</i>	Leaves/ Aqueous	Alkaloids, lycorine, crinamine	Okpo and Adeyemi, 2002
<i>Curcuma longa</i>	Rhizome	Tumerones, curcuminoids	Ammon and Wahl, 1991
<i>Elaeocarpus sphaericus</i>	Fruits/ Aqueous, pet-ether, benzene, acetone and ethanol	Glycoside, steroids, alkaloid, flavanoids	Singh <i>et al.</i> , 2000
<i>Gleditsia sinensis</i>	Fruits/ Ethanolic	Saponins	Dai <i>et al.</i> , 2002
<i>Impatiens textori</i>	Flowers/ Ethanolic	Apigenin, luteolin, chrysoeriol	Ishiguro <i>et al.</i> , 2000
<i>Inula mcmosa</i>	Roots/ Alcoholic	Inulolide-a new sesquiterpene, lactone	Srivastava <i>et al.</i> , 1999
<i>Magnolia officinalis</i>	Bark/ Aqueous	Honokiol, magnolol	Shin <i>et al.</i> , 2001b
<i>Mentha piperita</i>	Leaves	Flavanoidal glycosides	Inoue <i>et al.</i> , 2002
<i>Ocimum sanctum</i>	Leaves/ Aqueous	Myrcenol, nerol, eugenol	Sen, 1993
<i>Picrorrhiza kurroa</i>	Roots	Androsin	Stuppner <i>et al.</i> , 1991
<i>Siegesbeckia glabrescence</i>	Aerial parts/ Aqueous		Kang <i>et al.</i> , 1997
<i>Solanum xanthocarpum</i>	Roots/ Alkaloidal fraction	Solasodine	Chitravanshi <i>et al.</i> , 1990
<i>Striga orobanchioids</i>	Aerial parts/ Ethanolic		Harish <i>et al.</i> , 2001
<i>Tephrosia purpurea</i>	Aerial parts/ Ethanolic extract	Flavanoids, tephrosin	Gokhale <i>et al.</i> , 2000
<i>Terminalia chebula</i>	Fruits/ Aqueous	Ellagic acid, tannins, chebulagic acid	Shin <i>et al.</i> , 2001a
<i>Tinospora cordifolia</i>	Stem/ Aqueous	Tinosporin	Nayampalli <i>et al.</i> , 1986
<i>Tylophora asthmatica</i>	Leaves/ Alkaloidal	Tylophorine	Geetha <i>et al.</i> , 1981
<i>Vitex negundo</i>	Leaves/ Ethanolic	Casticin, isoorientin, chrysofenol D, luteolin	Nair <i>et al.</i> , 1994

**Anti-spasmodics**

*Aegle marmelos*, *Asiasarum sieboldi*, *Asystasia gangetica*,

*Bacopa monniera*, *Belamcanda chinensis*, *Cissampelos glaberrina*, *Clerodendron serratum*, *Cnidium monnieri*,

**Table 3. Anti-allergics**

Name of plant	Part used/ Extract/Fraction	Major chemical constituent (s)	References
<i>Adhatoda vasica</i>	Leaves/Methanolic	Vasicinol, vasicine	Muller <i>et al.</i> , 1993
<i>Albizia lebeck</i>	Stem bark/ Aqueous	Saponins	Baruah <i>et al.</i> , 1997
<i>Alisma orientale</i>	Rhizomes/ Aqueous, methanolic	Alisol B monoacetate, alismaketones-B 23-acetate and -C 23-acetate	Kubo <i>et al.</i> , 1997
<i>Aquillaria agallocha</i>	Stem/ Aqueous extract	Triterpenoids	Kim <i>et al.</i> , 1997
<i>Asiasarum sieboldi</i>	Roots/ Methanolic	Methyleugenol, $\gamma$ -asarone, elemicin, asarinin	Hashimoto <i>et al.</i> , 1994
<i>Camellia sinensis</i>	Leaves	flavonoids	Suzuki <i>et al.</i> , 2000
<i>Centipeda minima</i>	Aerial parts	flavonoids, pseudoguaienolide, sesquiterpene lactones	Wu <i>et al.</i> , 1985
<i>Citrus unshiu</i>	Peels	flavonoids	Kim <i>et al.</i> , 1999
<i>Cnidium monnieri</i>	Fruits/ Ethanolic	osthol	Matsuda <i>et al.</i> , 2002
<i>Crinum glaucum</i>	Leaves/ Aqueous	Alkaloids, lycorine, crinamine	Okpo and Adeyemi, 2002
<i>Dalbergia odorifera</i>	Heart Wood	Flavanoids, tannins	Chan <i>et al.</i> , 1998
<i>Desmodium adscendens</i>	Aqueous	Triterpenoid saponin	Addy, 1989
<i>Galphimia glauca</i>	Aerial/ Alcoholic extract /Ethyl-acetate	Tetragalloylquinic acid, quercetin	Neszmelyi <i>et al.</i> , 1993
<i>Ginkgo biloba</i>	Leaves	Ginkgolides	Touvay <i>et al.</i> , 1985
<i>Gleditsia sinensis</i>	Fruits/ Ethanolic	Saponins	Dai <i>et al.</i> , 2002
<i>Hydrangea macrophylla</i>	Leaves	Glycosides	Matsuda <i>et al.</i> , 1999
<i>Inula racemosa</i>	Roots/ Alcoholic	Inulolide-a new Sesquiterpene lactone	Srivastava <i>et al.</i> , 1999
<i>Magnolia officinalis</i>	Bark/ Aqueous	Honokiol, magnolol	Shin <i>et al.</i> , 2001b
<i>Sarcostemma brevistigma</i>	Twigs/ Alkaloidal fraction	Bregenin	Saraf and Patwardhan, 1988a
<i>Siegesbeckia glabrescence</i>	Aerial parts/ Aqueous		Kang <i>et al.</i> , 1997
<i>Solanum xanthocarpum</i>	Roots/ Alkaloidal fraction	Solasodine	Chitravanshi <i>et al.</i> , 1990
<i>Terminalia chebula</i>	Fruits/ Aqueous	Ellagic acid, Tannins chebulagic acid	Shin <i>et al.</i> , 2001a
<i>Vitex negundo</i>	Leaves/ Ethanolic	Casticin, isoorientin chrysophenol D, luteolin	Nair and Saraf, 1995

*Coleus forskohlii*, *Crinum glaucum*, *Drymis winteri*, *Ferula ovina*, *Ferula sinica*, *Pavetta crassipes*, *Saussurea leppa*, *Striga orobanchioids*, *Thymus vulgaris*, *Tylophora asthmatica* (Table 5).

#### Lipoxygenase inhibitors

*Allium cepa*, *Boswellia serrata*, *Coleus forskohlii*, *Lonicera japonica* (Table 6).

#### Platelet activating factor (PAF) inhibitors

*Allium cepa*, *Galphimia glauca*, *Impatiens textori*, *Picrorrhiza kurroa* (Table 6).

#### Cyclooxygenase inhibitor

*Allium cepa* (Table 6).

### SOME COMMONLY USED ANTI-ASTHMATIC HERBS

#### *Adhatoda vasica*

The medicinal properties of *Adhatoda vasica* Nees (natural order: Acanthaceae), called *Vasa* or *Vasaka* has been recommended by Ayurvedic physicians for the management of various types of respiratory disorders. The leaves of the plant were found to

**Table 4.** Anti-inflammatory agents

Name of plant	Part used/ Extract/Fraction	Major chemical constituent (s)	References
<i>Asystasia gangetica</i>	Leaves/ Methanolic, ethyl acetate	Isoflavone glycoside, dalhorinin	Akah <i>et al.</i> , 2003
<i>Calotropis procera</i>	Latex	$\alpha$ -amyrin, $\beta$ -amyrin calotropin (triterpenoid)	Kumar and Basu, 1994
<i>Curcuma longa</i>	Rhizomes	Tumerones, curcuminoids	Ammon and Wahl, 1991
<i>Dalbergia odorifera</i>	Heart Wood	Flavanoids, tannins	Chan <i>et al.</i> , 1998
<i>Elaeocarpus sphaericus</i>	Fruits/ Aqueous, pet-ether, benzene, acetone and ethanol	Glycoside, steroids, alkaloid, flavanoids	Singh <i>et al.</i> , 2000
<i>Ocimum sanctum</i>	Leaves/ Aqueous	Myrcenol, nerol, eugenol	Singh and Agrawal, 1991
<i>Pavetta crassipes</i>	Leaves/ Aqueous	Flavanoids, tannins, anthraquinones	Amos <i>et al.</i> , 1998
<i>Tylophora asthmatica</i>	Leaves/ Alkaloidal	Tylophorine	Manez, 1990

**Table 5.** Anti-spasmodic agents

Name of plant	Part used/Extract/Fraction	Major chemical constituent (s)	References
<i>Aegle marmelos</i>	Leaves/ Ethanolic	Aegelin, aegelemine, aegeline	Arul <i>et al.</i> , 2004
<i>Asiasarum sieboldii</i>	Roots/ Methanolic	Methyleugenol, $\gamma$ -asarone, elemicin, asarinin	Hashimoto <i>et al.</i> , 1994
<i>Asystasia gangetica</i>	Leaves/ Methanolic, ethyl acetate	Isoflavone glycoside, dalhorinin	Akah <i>et al.</i> , 2003
<i>Bacopa monniera</i>	Leaves/ Ethanolic	Bacosides, alkaloids, glycosides	Dar and Channa, 1997; Channa <i>et al.</i> , 2003
<i>Belamcanda chinensis</i>	Leaves/ Ethanolic	Tectorigenin	Singh and Agrawal, 1990
<i>Cissampelos glaberrima</i>	Leaves, root bark/ Aqueous	Warifiteine, $\alpha$ -bisbenzylisoquinoline alkaloid	Thomas <i>et al.</i> , 1995; Cortes <i>et al.</i> , 1995
<i>Clerodendron serratum</i>	Stem bark/ Aqueous	Phenolic glycoside	Gupta, 1968
<i>Cnidium monnieri</i>	Fruits/ Ethanolic	Osthol	Chen <i>et al.</i> , 1988
<i>Coleus forskohlii</i>	Roots	Forskolin (diterpenoid)	Marone <i>et al.</i> , 1987
<i>Crinum glaucum</i>	Leaves/ Aqueous	Alkaloids, lycorine, crinamine	Okpo and Adeyemi, 2002
<i>Drymis winteri</i>	Bark	Terpene	El-Sayah <i>et al.</i> , 1998
<i>Ferula ovina</i>	Aerial parts/ Ethanolic	Carvacrol, $\alpha$ -pinene, geranyl isovalerate and geranyl propionate	Khalil <i>et al.</i> , 1990
<i>Ferula sinica</i>	Roots/ Ethanolic		Aqel <i>et al.</i> , 1991a
<i>Pavetta crassipes</i>	Leaves/ Aqueous	Flavanoids, tannins, anthraquinones	Amos <i>et al.</i> , 1998
<i>Saussurea leppa</i>	Alkaloidal fraction	Sesquiterpene lactone, terpenoids	Dutta <i>et al.</i> , 1968
<i>Striga orobanchioids</i>	Aerial parts/ Ethanolic		Harish <i>et al.</i> , 2001
<i>Thymus vulgaris</i>	Ethanolic	Flavanones	Meister <i>et al.</i> , 1999
<i>Tylophora asthmatica</i>	Leaves/ Alkaloidal	Tylophorine	Haranath <i>et al.</i> , 1975; Udapa <i>et al.</i> , 1991

contain an essential oil and the quinazoline alkaloids vasicine, vasicinone and deoxyvasicine, which found to possess respiratory stimulant activity (Amin and Mehta, 1959). Of the two alkaloids, vasicinone was found to be more potent than vasicine, with potential

anti-asthmatic activity comparable to that of disodium cromoglycate (Atal and Kapur, 1982). Subacute toxicity of the alcoholic extract of leaves revealed that LD<sub>50</sub> of the extract by i.p. route was 581 mg/kg. The acute toxicity studies showed that extract was

**Table 6.** Miscellaneous agents

Name of plant	Part used/Extract/Fraction	Major chemical constituent (s)	References
Lipoxygenase inhibitors			
<i>Allium cepa</i>	Bulbs/Juice	$\alpha$ and $\beta$ unsaturated thiosulphinates	Bayer <i>et al.</i> , 1989
<i>Boswellia serrata</i>	Gum resin/Ethanollic extract	Boswellic acid	Ammon <i>et al.</i> , 1991
<i>Coleus forskohlii</i>	Roots	Forskolin (diterpenoid)	Marone <i>et al.</i> , 1987
Platelet Activating Factor (PAF) inhibitors			
<i>Allium cepa</i>	Bulbs/Juice	$\alpha$ and $\beta$ unsaturated thiosulphinates	Dorsch <i>et al.</i> , 1987
<i>Galphimia glauca</i>	Aerial/ Alcoholic extract/ Ethyl-acetate	Tetragalloylquinic acid, quercetin	Neszmelyi <i>et al.</i> , 1993
<i>Impatiens textori</i>	Flowers/Ethanollic	Apigenin, luteolin, chrysoeriol	Ueda <i>et al.</i> , 2003
<i>Picrorrhiza kurroa</i>	Roots	Androsin	Stuppner <i>et al.</i> , 1991
Cyclooxygenase inhibitor			
<i>Allium cepa</i>	Bulbs/Juice	$\alpha$ and $\beta$ unsaturated thiosulphinates	Bayer <i>et al.</i> , 1989

not lethal up to the dose of 100 mg/kg, i.p. and up to 4 g/kg (Rao and Krishnaiah, 1981). The LD<sub>50</sub> of alcoholic extract of the aerial parts of the plant is reported to be more than 1,000 mg/kg, i.p. in mice (Bhakuni *et al.*, 1990).

#### *Albizzia lebeck*

*Albizzia lebeck* has been used by Ayurvedic physicians for centuries in the management of asthma. The effect of decoction of the bark and flower were studied for its anti-asthmatic and anti-anaphylactic activity. The decoction protected the guinea pig against histamine and acetylcholine-induced bronchospasm (Tripathi and Das, 1977). The decoction of the bark of *Albizzia lebeck* was also studied on degranulation rate of sensitized peritoneal mast cells of albino rats when challenged with antigen (horse serum) and triple vaccine was used as adjuvant. Disodium cromoglycate (DCG) and prednisolone were used for comparison. Studies revealed the significant cromoglycate like action on the mast cells, which has been attributed to the heat-sable and water-soluble saponins present in the plant (Tripathi *et al.*, 1979). Crude extract of seeds and a pure saponin fraction of *Albizzia* have also been studied on the mast cells in the mesentery and peritoneal fluid of rats subject to anaphylaxis (Johri *et al.*, 1985). The Maximum

Tolerated Dose (MTD) of 50% ethanolic extracts of the root, the pods and stem bark was 25, 50 and 100 mg/kg i.p. in mice (Dhar *et al.*, 1968).

#### *Ammi visnaga*

*Ammi visnaga*, conventional anti-asthmatic compounds, such as sodium cromolyn and sodium cromoglycate, were developed from analogs of the naturally occurring furanochromone khellin (visamin), found in this Asian plant. Other furanochromones, such as visnagin, khellol and khellinol have also been identified in the extracts of *Ammi visnaga*. Khellin has been found to be an effective smooth muscle relaxant with an oral LD<sub>50</sub> of 80 mg/kg in rats. Controlled clinical studies have verified the anti-allergic action of sodium cromolyn, which is currently used in the treatment of allergic rhinitis, asthma and allergic gastrointestinal reactions (Johri *et al.*, 1985). The LD<sub>50</sub> of aqueous extract of *Ammi visnaga* of intraperitoneal (i.p.) and oral administration was 3.6 and 10.1 g/kg, respectively (Juoad *et al.*, 2002).

#### *Boswellia serrata*

The gum resin of *Boswellia serrata*, known in Indian Ayurvedic system of medicine as Salai guggal, contains boswellic acid. It specifically inhibits leukotriene biosynthesis by inhibiting the activity

of the enzymes, which leads to their formation. It also proved to be the most potent inhibitors of the classical component pathway of the inflammatory response. Boswellic acids also decrease the activity of human leukocyte elastase (HLE), which may be involved in the pathogenesis of emphysema. Boswellic acids are therefore effective in the prevention and or control of inflammatory processes, which are typically characterized by increased leukotriene formation (Safayhi *et al.*, 1997). Boswellia specifically blocks the synthesis of pro-inflammatory 5-lipoxygenase products, including leukotriene<sub>B<sub>4</sub></sub> (Ammon *et al.*, 1991), which cause bronchoconstriction, chemotaxis, and increased vascular permeability. Therefore Boswellic acid might be used for their anti-allergic/anti-asthmatic activity (Ammon *et al.*, 1991). LD<sub>50</sub> of alcoholic extract of gum resin was more than 2 g/kg, p.o. and i.p. in mice (Atal *et al.*, 1981; Singh and Atal, 1986). The Maximum Tolerated Dose (MTD) values of the root, fruit and stem extracts were 50, 500 and 250 mg/kg i.p. respectively in mice (Dhar *et al.*, 1968).

#### ***Clerodendron serratum***

*Clerodendron serratum* is widely used to alleviate the symptoms of respiratory conditions, including asthma. The root bark yields a phenolic glycoside (Vasavada *et al.*, 1967) and about 10% D-mannitol (Kirtikar and Basu, 1993). A sterol glycoside mixture was isolated. Hydrolysis of the crude sapogenin mixture of the bark yielded three major triterpenoid constituents-oleonolic acid, queretoroic acid and serratagenic acid. Gamma-sitosterol has also been isolated (Gupta and Gupta, 1967). It blocked the histamine-induced contractions of tracheal preparations from guinea pig without affecting the response to acetylcholine (Vasavada *et al.*, 1967). It is reported that the continuous daily administration of the plant extract to sensitized guinea pig, gradually developed protection against anaphylaxis. The saponin also disrupted rat peritoneal mast cells and blocked the effect of horse serum antigen. Saponins from the root caused disruption of cells

of rat mesentery in a dose related manner (up to a dose of 40 µg), and the maximum disruption effect was exerted in 30 min (Gupta *et al.*, 1971).

#### ***Curcuma longa***

*Curcuma longa*, by virtue of its antioxidant properties is an effective anti-asthmatic agent. It has been employed by Ayurvedic practitioners since ancient times in the treatment of respiratory disorders. The active ingredients, the curcuminoids, are potent inhibitors of inflammatory prostaglandins. The overall anti-inflammatory action of curcuminoids is also related to their well-known antioxidant properties. For example, curcumin inhibited lipid peroxidation, a phenomenon associated with antioxidant as well as anti-inflammatory activities. Toxicity studies of *Curcuma longa* revealed that acute doses of 0.5, 1.0, and 3 g/kg body weight and the chronic doses of 100 mg/kg/day of ethanolic extract were found to be non-toxic (Quereshi *et al.*, 1998).

#### ***Ephedra sinica* (Ma Huang)**

*Ephedra sinica*, a native plant species of China is the original source of the alkaloid ephedrine. Ephedrine stimulates the sympathetic nervous system, and thereby helps in the management of allergic conditions. The compound also helps to relieve the bronchial spasm that underlies conditions such as asthma and emphysema through this effect. As ephedrine use is now restricted in several countries, alternatives such as *Citrus aurantium* (containing synephrine) are now being explored in the management of respiratory conditions. Other ancillary alternative phytonutrients useful in asthma include licorice which has been used as an expectorant. Phytonutrients are often included in anti-asthmatic formulations, with antioxidants such as N-acetylcysteine which prevent mucus build-up and inhibit free radical mediated disease processes. Oral LD<sub>50</sub> of d-pseudoephedrine and 1-ephedrine was 1,550 (1,360 - 1,767) mg/kg and 1,400 (1,102 - 1,778) mg/kg, respectively. LD<sub>50</sub> of d-pseudoephedrine and 1-ephedrine given

intraperitoneally was 245 (229 - 262) mg/kg and 300 (259 - 348) mg/kg, respectively (Akiba *et al.*, 1979).

### *Picrorhiza kurroa*

*Picrorhiza kurroa* Royle is a perennial herb that grows in the Himalayas in Asia, at altitudes of 9,000 - 15,000 feet above sea level. It belongs to the Natural Order *Scrophulariaceae*. The underground parts of this plant have been used in the traditional Indian systems of medicine since ancient times to treat liver troubles and bronchial problems (Kirtikar and Basu, 1993). Several biologically active principles, particularly glycosides have been identified in extracts obtained from *Picrorhiza kurroa*. Of these a mixture of the iridoid glycosides picroside I and kutkoside has been found to be an efficient liver protectant. "Androsin", a phenolic glycoside isolated from *Picrorhiza kurroa*, has been attributed with anti-asthmatic properties (Dorsch *et al.*, 1991).

The authors suggest that androsin may act by depressing the activity of PAF which plays a major role in the pathogenesis of bronchial asthma. PAF has been shown to provoke long-lasting inflammatory responses in the lungs. This leads to bronchial hyperactivity and subsequent bronchial obstruction (Dorsch *et al.*, 1991). Another study suggests that *Picrorhiza kurroa* extracts possess anti-allergic activity, probably mediated through mast cell stabilizing activity (Mahajani and Kulkarni, 1977).

### *Tylophora asthmatica* (syn. *Tylophora indica*)

The medicinal properties of the plant *Tylophora asthmatica* have been known since ancient times. Powder from the dried leaves, root powder, and decoction of the leaves or infusion of the root bark have been used traditionally in the treatment of respiratory affections such as chronic bronchitis and asthma (Nadkarni, 1976). Preparations containing dried, powdered plant material are available for the treatment of bronchial asthma and tropical eosinophilia. The anti-asthmatic activity of the plant is attributed to the presence of

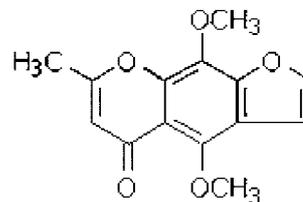


Fig. 1. Chemical structure of khellin.

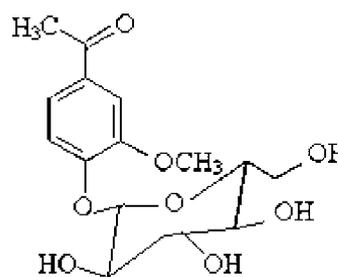


Fig. 2. Chemical structure of androsin.

phenanthroindolizidine alkaloids, which has been isolated from the aerial parts of the plant (Ali and Bhutani, 1989). A water extract of the plant showed anti-anaphylactic effect, leucopenia and inhibition of Schulz-Dale's reaction in experimental animals. The extract also showed brief nonspecific anti-spasmodic action in isolated tissues of g. pig ileum, rabbit duodenum, frog's rectus and rat stomach. The mode of action of the plant may be cell-mediated immunity (Haranath and Shyamalakumari, 1975). The plant extracts were found to produce significant anti-inflammatory effects in rats (Manez 1990). Immunosuppressive and anti-inflammatory effects of *Tylophora asthmatica* are due to increased secretion of corticosteroids by adrenal cortex (Udupa, 1991). *Tylophora asthmatica* also produced significant improvement in lung functions, when the effect of the plant was studied on the patients of bronchial asthma (Gore, 1980). Preliminary studies on animals have found *tylophora* extracts to be toxic only in extremely high doses; these extracts were apparently safe in the far smaller doses needed to produce a therapeutic effect (Dikshith *et al.*, 1990).

## CONCLUSIONS

Herbal approaches have regained their popularity, with their efficacy and safety aspects being supported by controlled clinical studies. The herbal approach have offered effective mast cell stabilizers like sodium cromolyn and sodium cromoglycate developed from khellin and anti-leukotriene product - boswellic acids. Ongoing research worldwide has provided valuable clues regarding the precise mechanism of action of these herbal alternatives and these herbs, therefore, have multi-faceted roles to play in the management of asthma. Some herbal alternatives employed in these traditions are proven to provide symptomatic relief and assist in the inhibition of disease development as well. Thus, these plants can be used to obtain a polyherbal formulation which contains various herbs acting at particular sites of the pathophysiological cascade of asthma for prophylaxis as well as for the treatment of asthma. Further, different formulations can be prepared which can be used in different types of respiratory disorders including different types of asthma.

## REFERENCES

- Acharya SB, Yanpallewar SU, Singh RK. (2003) A preliminary study on the effect of *Azadirachta indica* on bronchial smooth muscles and mast cells. *J. Nat. Remed.* **3**, 78-82.
- Addy ME. (1989) Several chromatographically distinctive fractions of *Desmodium adscendens* inhibit smooth muscle contractions. *Int. J. Crude Drug Res.* **27**, 81-91.
- Agrawal BB, Mehta AA. (2005) Phyto-pharmacological investigation of *Moringa oleifera* and *Achyranthus aspera* for their anti-asthmatic activity. Ph.D. thesis, Gujarat University.
- Akah PA, Ezike AC, Nwafor SV, Okoli CO, Enwerem NM. (2003) Evaluation of the anti-asthmatic property of *Asystasia gangetica* leaf extracts. *J. Ethnopharmacol.* **89**, 25-36.
- Akiba K, Onodera K, Kisara K, Fujikura H. (1979) Interaction of d-pseudoephedrine with water soluble extracts of *Platycodi Radix* on acute toxicity. *Nippon Yakurigaku Zasshi* **75**, 201-206.
- Ali M, Bhutani KK. (1989) Alkaloids from *Tylophora indica*. *Phytochemistry* **28**, 3513-3517.
- Amin AH, Mehta DR. (1959) Bronchodilator alkaloid from *Adhatoda vasica*. *Nature* **184**, 1317.
- Ammon HP, Mack T, Singh GB, Safayhi H. (1991) Inhibition of leukotriene B<sub>4</sub> formation in rat peritoneal neutrophils by an ethanolic extract of the gum resin exudates of *Boswellia serrata*. *Planta med.* **57**, 203-207.
- Ammon HP, Wahl MA. (1991) Pharmacology of *Curcuma longa*. *Planta Med.* **57**, 1-7.
- Amos S, Gamaniel K, Akah P, Wambebe C. (1998) Anti-inflammatory and muscle relaxant effect of aqueous extract of *Pavetta crassipes* leaves. *Fitoterapia* **69**, 425-29.
- Aqel MB. (1991) Relexant effect of the volatile oil of *Romarinus officinalis* on tracheal smooth muscle. *J. Ethnopharmacol.* **33**, 57-62.
- Arul V, Miyazaki S, Dhananjayan R. (2004) Mechanisms of the contractile effect of the alcoholic extract of *Aegle marmelos* Corr. on isolated guinea pig ileum and tracheal chain. *Phytomedicine* **11**, 679-683.
- Atal CK, Gupta OP, Singh GB. (1981). Salai guggal: A promising anti-arthritis and anti-hyperlipidaemic agent. *Br. J. Pharmacol.* **74**, 113-119.
- Atal CK, Kapur BM. (1982) *Cultivation and Utilization of Aromatic Plants*, pp. 155, Regional Research Laboratory, Council of Scientific and Industrial Research, Jammu-Tawi, India.
- Baruah CC, Gupta PP, Patnaik GK, Nath A, Kulshreshtha DK, Dhawan BN. (1997) Anti-allergic and mast cell stabilizing activity of *Albizia lebeck*. *Indian Veterin. Med. J.* **21**, 127-132.
- Bayer T, Breu W, Seligmann O, Wray V, Wagner H. (1989) Biologically active thiosulphinates and a sulphanyl disulphides from *Allium cepa*. *Phytochemistry* **28**, 2373-2377.
- Bhakuni DS, Goel AK, Jain S, Mehrotra BN, Srimal RC. (1990) Screening of Indian plants for biological activity. Part XIV. *Indian J. Exp. Biol.* **28**, 619-637.
- Campos MG, Toxqui E, Tortoriello J, Oropeza MV, Ponce H, Vargas MH, Montano LM. (2001) *Galphimia glauca* organic fraction antagonized LTD (4)-induced contraction in guinea pig airways. *J. Ethnopharmacol.* **74**, 7-15.

- Chan SC, Chang YS, Wang JP, Chen SC, Kuo SC. (1998) Three new flavonoids and anti-allergic, anti-inflammatory constituents from the heartwood of *Dalbergia odorifera*. *Planta Med.* **64**, 153-158.
- Channa S, Dar A, Yaqoob M, Anjum S, Sultani Z, Atta-ur-Rahman. (2003) Broncho-vasodilatory activity of fractions and pure constituents isolated from *Bacopa monniera*. *J. Ethnopharmacol.* **86**, 27-35.
- Charak Samhita. (1949) Shri Gulab Kunverba Ayurvedic Society, Jamnagar, Ayurvedic Mudranalaya, Jamnagar, Vol. IV, 1952-2032.
- Chen ZC, Duan XB, Liu KR. (1988) The anti-allergic activity of osthol extracted from the fruits of *Cnidium monnieri* (L.) Cusson. *Yao Xue Xue Bao* **23**, 96-99.
- Chitravanshi VC, Gupta PP, Kulshrestha DK, Kar K, Dhawan BN. (1990). Anti-allergic activity of *Solanum xanthocarpum*. *Indian J. Pharmacol.* **22**, 23-24.
- Cortes SF, Alencar JL, Thomas G, Filho JMB. (1995) Spasmolytic action of warifteine, a bisbenzylisoquinoline alkaloid isolated from the root bark of *Cissampelos sympodialis* Eichl. *Phytother. Res.* **9**, 579-583.
- Dai Y, Chan YP, Chu LM, Bu PP. (2002) Antiallergic and anti-inflammatory properties of the ethanolic extract from *Gleditsia sinensis*. *Biol. Pharm. Bull.* **25**, 1179-1182.
- Dar A, Channa S. (1997) Relaxant effect of ethanol extract of *Bacopa monniera* on trachea, pulmonary and aorta from rabbit and guinea pig. *Phytother. Res.* **11**, 323-325.
- Dhar ML, Dhar MM, Dhawan BN, Mehrotra BN, Ray C. (1968) Screening of Indian plants for biological activity. Part I. *Indian J. Exp. Biol.* **6**, 232-247.
- Dhawan K, Kumar S, Sharma A. (2003) Anti-asthmatic activity of the methanol extract of leaves of *Passiflora incarnata*. *Phytother. Res.* **17**, 821-822.
- Dikshith TS, Raizada RB, Mulchandani NB. (1990) Toxicity of pure alkaloid of *Tylophora asthamatica* in male rat. *Indian J. Exp. Biol.* **28**, 208-212.
- Dorsch W, Ettl M, Hein G, Scheftner P, Weber J, Bayer T, Wagner H. (1987) Anti-asthmatic effects of onions. Inhibition of platelet activating factor induced bronchial obstruction by onion oils. *Int. Arch. Allergy Appl. Immunol.* **82**, 535-536.
- Dorsch W, Stuppner H, Wagner H, Gropp M, Demoulin S, Ring J. (1991) Anti-asthmatic effect of *Picrorhiza kurroa*: androsin prevents allergen and PAF induced bronchial obstruction in guinea pigs. *Int. Arch. Allergy Appl. Immunol.* **95**, 128-133.
- Dutta NK, Sastry M, and Tamhane RG. (1968) Pharmacological actions of an alkaloidal fraction isolated from *Saussurea leppa* (Clarke). *Curr. Sci.* **37**, 550-551.
- Fireman P. (2003) Understanding asthma pathophysiology. *Allergy Asthma Proc.* **24**, 79-83.
- Geetha VS, Viswanathan S, Kameswaran L. (1981) Comparison of total alkaloids of *Tylophora indica* and disodium cromoglycate on mast cell stabilization. *Indian J. Pharmacol.* **13**, 199-201.
- Gokhale AB, Dikshit VJ, Damre AS, Kulkarni KR, Saraf MN. (2000) Influence of ethanolic extract of *Tephrosia purpurea* Linn. on mast cells and erythrocytes membrane integrity. *Indian J. Exp. Biol.* **38**, 837-840.
- Gore KV, Rao K, Guruswamy MN. (1980) Physiological studies with *Tylophora asthamatica* in bronchial asthma. *Indian J. Med. Res.* **71**, 144-148.
- Gupta SS, and Gupta MK. (1967) Effect of *Solanum xanthocarpum* and *Clerodendron serratum* on histamine release from tissues. *Indian J. Med. Sci.* **21**, 795-799.
- Gupta SS. (1968) Development of anti-histaminic and anti-allergic activity after prolonged administration of a plant saponin from *Clerodendron serratum*. *J. Pharm. Pharmacol.* **20**, 801-802.
- Gupta SS. (1971) Effect of *Clerodendron serratum* on mast cells of rat mesentery. *Indian J. Med. Sci.* **25**, 29.
- Gupta SS, Tripathi RM. (1973) Effect of chronic treatment of the saponin of *Clerodendron serratum* on disruption of mesenteric mast cells of rats. *Aspects Allergy Appl. Immunol.* **6**, 177-188.
- Gupta SS. (1974) Some observations on the anti-asthmatic effect of the saponins of *Gardenia latifolia*. *Aspects Allergy Appl. Immunol.* **7**, 198-204.
- Haranath PSRK, Shyamalakumari S. (1975) Experimental study on the mode of action of *Tylophora asthamatica* in bronchial asthma. *Indian J. Med. Res.* **63**, 661-670.
- Hashimoto K, Yanagisawa T, Okui Y, Ikeya Y, Maruno M, Fujita T. (1994) Studies on anti-allergic components in the roots of *Asiasanum sieboldi*. *Planta Med.* **60**, 124-127.
- Inoue T, Sugimoto Y, Masuda H, Kamei C. (2002) Anti-allergic effect of flavonoids obtained from *Mentha piperita* L. *Biol. Pharm. Bull.* **25**, 256-259.
- Ishiguro K, Ueda Y, Iwaoka E, Oku H. (2000). Antiallergic and antipruritic effect of *Impatiens textori*. *Phytomedicine* **7**, 94-97.

- Johri RK, Zutshi U, Kameshwaran L, Atal CK. (1985) Effect of quercetin and *Albizia saponins* on rat mast cell. *Indian J. Physiol. Pharmacol.* **29**, 43-46.
- Jouad H, Maghrani M, Eddouks M. (2002) Hypoglycemic effect of aqueous extract of *Ammi visnaga* in normal and streptozotocin-induced diabetic rats. *J. Herb Pharmacother.* **2**, 19-29.
- Kanno M, Shibano T, Takido M, Kitanaka S. (1999) Anti-allergic agent from natural sources. 2. structures and leukotriene release-inhibitory effect of torososide B and torosachryson 8-O-6-malonyl beta gentiobioside from *Cassia torosa* Cav. *Chem. Pharm. Bull.* **47**, 915-918.
- Khalil SA, Aqel M, Afifi F, Eisawi DA. (1990) Effect of an aqueous extract of *Fenula ovina* on rabbit and guinea pig smooth muscle. *J. Ethnopharmacol.* **30**, 35-42.
- Kim YC, Lee EH, Lee YM, Kim HK, Song BK, Lee EJ, Kim HM. (1997) Effect of the aqueous extract of *Aquillaria agallocha* stem on the immediate hypersensitivity reactions. *J. Ethnopharmacol.* **58**, 31-38.
- Kim DK, Lee KT, Eun JS, Zee OP, Lim JP, Eum SS, Kim SH, Shin TY. (1999) Anti-allergic components from peels of *Citrus unshiu*. *Arch. Pharm Res.* **22**(6), 642-645.
- Kirtikar KR, Basu BD. (1993) *Indian Medicinal Plants*, Vol. III, pp. 194, Periodical Experts Book Agency, Delhi, India.
- Kitanaka S, Nakayama T, Shibano T, Ohkoshi E, Takido M. (1998) Anti-allergic agent from natural sources, structures and inhibitory effect of histamine release of naphthopyrone glycosides from seeds of *Cassia obtusifolia* L. *Chem. Pharm. Bull.* **46**, 1650-1652.
- Kubo M, Matsuda H, Tomohiro N, Yoshikawa M. (1997) Studies on *Alismatis rhizome*; Anti-allergic effects of methanol extract and six terpene components from *Alismatis rhizoma* (dried rhizome of *Alisma orientale*). *Biol. Pharm. Bull.* **20**, 511-516.
- Kumar VL, Basu N. (1994) Anti-inflammatory activity of the latex of *Calotropis procera*. *J. Ethnopharmacol.* **44**, 123-125.
- Kumar DA, Ramu P. (2002) Effect of methanolic extract of *Benincasa hispida* against histamine and acetylcholine induced bronchospasm in guinea pigs. *Indian J. Pharmacol.* **34**, 365-366.
- Mahajani SS, Kulkarni RD. (1977) Effect of DSCG and *P. kurroa* root powder on sensitivity of guinea pigs to histamine and sympathomimetic amines. *Int. Arch. Allergy Appl. Immunol.* **53**, 137-144.
- Manez S, Alcaraz MJ, Paya M, Rios JL, Hancke JL. (1990) Selected extracts from medicinal plants as anti-inflammatory agents. *Planta Med.* **56**, 656.
- Marone G, Columbo M, Triggiani M, Cirillo R, Genovese A, Formisano S. (1987) Inhibition of IgE mediated release of histamine and peptide leukotriene from human basophils and mast cells by forskolin. *Biochem. Pharmacol.* **36**, 13-20.
- Matsuda H, Shimoda H, Yamahara J, Yoshikawa M. (1999) Effect of phylloolucin, hydrangenol, and their 8-O-glucosides, and thunberginols A and F from *Hydrangea macrophylla* var. *thunbergii* on passive cutaneous anaphylaxis reaction in rats. *Biol. Pharm. Bull.* **22**, 870-872.
- Matsuda H, Tomohiro N, Ido Y, Kubo M. (2002) Anti-allergic effects of *Cnidii monnieri* (dried fruits of *Cnidium monnieri*) and its major component, osthol. *Biol. Pharm. Bull.* **25**, 809-812.
- Meister A, Bernhard G, Chrisoffel V, Buschauer A. (1999) Antispasmodic activity of *Thymus vulgaris* extract on isolated g.pig trachea: discrimination between drug and ethanol effects. *Planta Med.* **65**, 512-516.
- Moran A, Carron R, Martin ML, San Roman L. (1989) Anti-asthmatic activity of *Artemisia caerulea* subsp. *gallica*. *Planta Med.* **55**, 351-353.
- Muller A, Antus S, Bittinger M, Dorsch W, Kaas A, Kreher B *et al.* (1993) Chemistry and pharmacology of the antiasthmatic plants *Galphimia glauca*, *Adhatoda vasica* and *Picorrhiza kurroa*. *Planta Med.* **59**, A586.
- Nadkarni AK. (1976) *Indian Materia Medica*, pp. 810, Popular Prakashan; Bombay.
- Nair AM, Tamhankar CP, Saraf MN. (1994) Studies on the mast cell stabilizing activity of *Vitex negundo* Linn. *Indian Drugs* **32**, 277-282.
- Nair AM, Saraf MN. (1995) Inhibition of antigen and compound 48/80 induced contractions of g.pig trachea by the ethanolic extract of the leaves of *Vitex negundo* Linn. *Indian J. Pharmacol.* **27**, 230-233.
- Nayampalli S, Desai NK, Ainapure SS. (1986) Anti-allergic properties of *Tinospora cordifolia* in animal models. *Indian J. Pharmacol.* **18**, 250-252.
- Neszmelyi A, Kreher B, Muller A, Dorsch W, Wagner H. (1993) Tetragalloylquinic acid, the major Antiasthmatic principle of *Galphimia glauca*. *Planta Med.* **59**, 164-167.

- Okpo SO, Adeyemi OO. (2002) The anti-allergic effects of *Crinum glaucum* aqueous extract. *Phytomedicine* **9**, 438-441.
- Palanichamy S, Amala Bhaskar E, Nagarajan S. (1991) Effect of *Cassia alata* leaf extract on mast cell stabilization. *Indian J. Pharmacol.* **23**, 189-191.
- Paliwa JK, Dwiwedi AK, Singh S. (2000) Pharmacokinetics and in-situ absorption studies of a new anti-allergic compound 73/602 in rats. *Int. J. Pharm.* **197**, 213-220.
- Puglisi L, Salvadori S, Gabrielli G, Pasargiklian R. (1988) Pharmacology of natural compounds. Smooth muscle relaxant activity induced by a *Ginkgo biloba* L. extract on guinea-pig trachea. *Pharmacol. Res. Comm.* **20**, 573-589.
- Qureshi S, Shah AH, Ageel AM. (1998) Toxicity studies on *Alpinia galanga* and *Curcuma longa*. *Indian J. Exp. Biol.* **36**, 675-679.
- Rao VSN, Krishnaiah KS. (1981) Pharmacological investigations on *Adhatoda vasica* Nees (vasaka). *Indian Vet. J.* **58**, 107-111.
- Safayhi H, Rall B, Sailer ER, Ammon HPT. (1997) Inhibition by Boswellic acids of human leukocyte elastase. *J. Pharmacol. Exp. Ther.* **281**, 460-463.
- Samiulla DS, Prashanth D, Amit A. (2001) Mast-cell stabilizing activity of *Bacopa monnieri*. *Fitoterapia* **72**, 284-285.
- Saraf MN, Patwardhan BK. (1988a) Pharmacological studies on *Sarcostemma brevistigma*. Part I Anti-allergic activity, *Indian Drugs* **26**, 49-53.
- Saraf MN, Patwardhan BK. (1988b) Pharmacological studies on *Sarcostemma brevistigma*. Part II Bronchodilator activity. *Indian Drugs*. **26**, 54-57.
- Schayck CP, Graafsma SJ, Visch MB, Dompeling E, Weel C, van Herwaarden CLA. (1990) Increase bronchial hyperresponsiveness after inhaling salbutamol during 1 year is not caused by subsensitization to salbutamol. *J. Allergy Clin. Immunol.* **86**, 793-800.
- Sen P. (1993) Therapeutic potential of Tulsi (*Ocimum sanctum*) from experience to fact. *Drug Views* **1**, 15-18.
- Shin TY, Jeong HJ, Kim DK, Kim SH, Lee JK, Chae BS, Kim JH, Kang HW, Lee CM. (2001a) Inhibitory action of water-soluble fraction of *Terminalia chebula* on systemic and local anaphylaxis. *J. Ethnopharmacol.* **74**, 133-140.
- Shin TY, Kim DK, Chae BS, Lee EJ. (2001b) Antiallergic action of *Magnolia officinalis* on immediate hypersensitivity reaction. *Arch. Pharm. Res.* **24**, 249-255.
- Shinde UA, Phadke AS, Kulkarni KR, Nair AM, Mungantiwar AA, Dikshit VJ, Saraf MN. (1999) Mast cell stabilizing and lipoxygenase inhibiting activity of *Cedrus deodara* (Roxb.) wood oil. *Indian J. Exp. Biol.* **37**, 258-261.
- Singh GB, Atal CK. (1986). Pharmacology of an extract of Salai guggal ex *Boswellia serrata*, a new non-steroidal anti-inflammatory agent. *Agents Actions* **18**, 407-412.
- Singh RK, Acharya SB, Bhattcharya SK. (2000) Pharmacological activity of *Elaeocarpus sphericus*. *Phytother. Res.* **14**, 36-39.
- Singh S, Agrawal SS. (1990) Broncho-relaxant activity of *Belamcanda chinensis*. *Indian J. Pharmacol.* **22**, 107-109.
- Singh S, Agrawal SS. (1991) Anti-asthmatic and anti-inflammatory activity of *Ocimum sanctum*. *Int. J. Pharm.* **29**, 306-310.
- Srivastava S, Gupta PP, Prasad R, Dixit KS, Palit G, Ali B, Mishra G, Saxena RC. (1999) Evaluation of anti-allergic activity (Type I hypersensitivity) of *Inula racemosa* in rats. *Indian J. Physiol. Pharmacol.* **43**, 235-241.
- Stuppner H, Dorsch W, Wagner H, Gropp M, Kepler P. (1991) Antiasthmatic effects of *Picorrhiza kurroa*: Inhibition of allergen and PAF induced bronchial obstruction in g.pigs by Androsin, Apocynine and structurally related compounds. *Planta Med.* **57**, A62.
- Suzuki M, Yoshino K, Yamamoto MM, Miyase T, Sano M. (2000) Inhibitory effect of Tea catechins and o-methylated derivatives of (-)-epigallocatechin-3-O-gallate on mouse type IV allergy. *J. Agric. Food Chem.* **48**, 5649-5653.
- Thomas G, Araujo CC, Agra MF, Diniz M. (1995) Preliminary studies on the hydroalcoholic extract of the root of *Cissampelos sympodialis* Eichl in guinea pig tracheal strips and bronchoalveolar leucocytes. *Phytother. Res.* **9**, 473-477.
- Thomas G, Araujo CC, Duarte JC, De souza DP. (1997) Bronchodilator activity of an aqueous fraction of an ethanol extract of the leaves of *Cissampelos sympodialis* Eichl. in the guinea pig. *Phytomedicine* **4**, 233-238.
- Touvy C, Eienne A, Braquet P. (1986) Inhibition of antigen induced lung anaphylaxis in the guinea pig by BN 52021 a new specific PAF-acether receptor antagonist isolated from *Ginkgo biloba*. *Agents Actions* **17**, 371-372.

- Tripathi RM, Das PK. (1977) Studies on anti-asthmatic and anti-anaphylactic activity of *Albizzia lebeck*. *Indian J. Pharmacol.* **9**, 189-194.
- Tripathi RM, Sen PC, Das PK. (1979) Studies on the mechanism of action of *Albizzia lebeck*, an Indian indigenous drug used in the treatment of atopic allergy. *J. Ethnopharmacol.* **1**, 385-386.
- Udapa AL, Udapa SL, Guruswamy MN. (1991) The possible site of anti-asthmatic action of *Tylophora asthmatica* on pituitary-adrenal axis in albino rats. *Planta Med.* **57**, 409-413.
- Ueda Y, Oku H, Inuma M, Ishiguro K. (2003) Effect on blood pressure decrease in response to PAF of *Impatiens textori*. *Biol. Pharm. Bull.* **26**, 1505-1507.
- Vasavada SA. (1967) Gamma sitosterol from *Clerodendron serratum*. *Bull. Calc. Sch. Trop. Med.* **15**, 61.
- Wang N, Yao X, Ishii R, Kitanaka S. (2001) Antiallergic agents from natural sources. structures and inhibitory effects on nitric oxide production and histamine release of five novel polyacetylene glucosides from *Bidens parviflora willd.* *Chem. Pharm. Bull.* **49**, 938-942.
- Wu JB, Chun YT, Ebizuka Y, Sankawa V. (1985) Biologically active constituents of *Centipeda minima*: Isolation of a new sesquiterpene lactones. *Chem. Pharm. Bull.* **33**, 4091-4094.